

INTERNATIONAL JOURNAL OF RESEARCH IN PHARMACEUTICAL SCIENCES

Published by Pharmascope Publications

Journal Home Page: <u>www.pharmascope.org/ijrps</u>

Phytochemical library of Caralluma genus

Sireesha M¹, Venkata Nadh R^{*2}, Suresh Babu K³, Sreenivasulu M¹

¹Department of Science and Humanities, Vignan's Foundation for Science, Technology and Research, Vadlamudi 522 213, India ²GITAM University, Bengaluru Campus, Karnataka – 561203, India ³Department of Chemistry, Mallareddy Engineering College, Hyderabad, India

Article History:	ABSTRACT
Received on: 13.07.2018 Revised on: 19.09.2018 Accepted on: 21.09.2018	<i>Caralluma</i> is one of the prominent genus out of 200 genera and 2500 species of Asclepiadaceae family. <i>Caralluma</i> genus belongs to Asclepiadaceae and is widely distributed in Asia countries, Africa, Arabian Peninsula, Canary Islands and Southeast Europe. In folkloric medicine, as well as in Unani and
Keywords:	Ayurvedic systems of medicine, the plants of <i>Caralluma</i> are being used for the treatment of diabetic patients and rheumatism. Tribals consider some
Asclepiadaceae, C <i>aralluma,</i> Medicinal plants, Pharmacological activities, Phytochemical library	them as food during famines and also as a part of the traditional medicinal system. In India and Pakistan, <i>Caralluma</i> species have been used as emergency foods for the last few centuries. As allopathic medicines possess toxic nature and side effects, the use of plant-based medicine is becoming popular. This lead to a sudden enhancement in the production of herbal drugs. At present <i>Caralluma</i> is gaining much importance from researchers because it possesses array immunostimulating activities due to presence flavonoids and saponins and pregnane glycosides as active components. The present article thoroughly reviewed about various phytochemicals present in different species of <i>Caralluma</i> . Tabulated a list of phytochemicals isolated by various researchers from different plants of the <i>Caralluma</i> genus. Further, listed out the species on which enthusiastic researchers of this field can extend their investigation.

* Corresponding Author

Name: Prof. R. Venkata Nadh Phone: +91-9902632733 Email: doctornadh@yahoo.co.in

ISSN: 0975-7538

DOI: <u>https://doi.org/10.26452/ijrps.v9i4.1655</u>

Production and Hosted by

Pharmascope.org © 2018 Pharmascope Publications. All rights reserved.

INTRODUCTION

Nature is the wonderful source of the compounds containing medicinal properties. Extraordinary chemical diversity can be found in millions of living species like animals, marine organisms, microorganisms and plants (Newman *et al.*, 2003; Bhanot *et al.*, 2011). The term natural product means it is

a chemical substance found in living organisms viz., marine organism, plants and fermentation of microbes (Baker et al., 2007). Natural products extraction and their uses are well known from ancient times since the discovery of fire (Chemat and Strube, 2015). Natural products are becoming a significant source for the development of new drugs and new chemical moieties. 60% of presently using anticancer agents is obtained from the plants/natural sources (Cragg et al., 2011). All the chemical compounds of any biological system can be classified into two broad categories. The first category is the primary metabolites and the second category is secondary metabolites. The chemical substances which are meant for growth and development of living body are called as primary metabolites. They include amino acids, carbohydrates, lipids and proteins. The group of chemical substances other than primary metabolites those existing in a biological system are called as secondary metabolites. Presence of secondary metabolites in

living body gives the ability to face the challenges in its survival, during the time of its interaction with the surroundings (Harborne, 1993).

Medicinal Plants

According to WHO, the medicinal plant can be defined as a plant in which one or more of its parts include substances containing therapeutic applications or compounds which are responsible for semi-synthesis of pharmaceuticals. Since olden days medicinal plants are standing as valuable sources for therapeutic compounds, many of existing drugs till today are derived from plant based natural products or their derivatives (Newman and Cragg, 2012). Use of plants for humankind is older as the begin of mankind. Nowadays medicinal plants are gaining much importance from people who are living in urban areas in treating infections like typhoid fever and gonorrhoea. This is due to the inefficiency of modern medicines and increased resistance shown by various bacteria towards antibiotics, as well as high cost of prescribed drugs in the maintenance of health (Smolinski et al., 2003; Van den Bogaard and Stobberingh, 2000). Unfortunately, all over the world modern health facilities available are not meeting the demands of the increased human population. Therefore it is demanding the further use of natural remedies in maintaining the health of humans. Various diseases like asthma, constipation, fever and hypertension were treated using traditional medicinal plants (Saganuwan, 2010).

Phytochemicals

The medically active components extracted from plants are also known as phytochemicals or phytoconstituents and were playing a vital role in protecting the plant from various diseases caused by pests and microbes (Doughari et al., 2009; Nweze et al., 2004). Phytochemical is a Greek word and it means plant-derived chemical. In recent times it is well known that phytochemicals are also playing a vital role in maintaining human health, particularly if they are taken as significant diet (Saxena et al., 2013). Phytochemicals are widely distributed in foods like herbs, spices, legumes, vegetables and fungi (Mathai, 2000). Phytochemicals exist in various parts of plants like whole grains, seeds, flowers, fruits, stems, bark, leaves and roots (Costa et al., 1999). Phytochemistry is one of the branches of chemistry that involves the study of the relationship between organic chemistry and natural products. The studies which are associated with phytochemistry are very significant and appropriate as it helps to gain the knowledge on a range of plantbased chemicals those are pharmacologically important (Temidayo, 2013). A number of phytochemicals were separated and characterized from

certain sources like fruits (apples and grapes), vegetables (onion and broccoli), spices (turmeric), beverages (red wine and green tea), and from many other sources (Doughari *et al.*, 2009; Doughari and Obidah, 2008).

Caralluma Genus

Caralluma genus belongs to Asclepiadaceae and is widely distributed in Asia (countries like Afghanistan, India, Iran, Pakistan and Sri Lanka), Africa, Arabian Peninsula, Canary Islands and Southeast Europe (Meve et al., 2004; Gilbert 1990). In folkloric medicine, as well as in Unani and Ayurvedic systems of medicine, the plants of Caralluma are being used for the treatment of diabetic patients and rheumatism (Ramesh et al., 1998). Tribals consider some of them as food during famines (Ahmad et al., 1989) and also as a part of the traditional medicinal system (Abdul-Aziz Al-Yahya et al., 2000). In India and Pakistan, Caralluma species have been used as emergency foods for the last few centuries (Gandhi, 1999). A spectrum of biological activities of Caralluma species can be expected due to the existence of pregnane glycosides, stigmasterol and other phytochemicals in them (Malladi et al., 2017; 2018; Suresh Babu et al., 2014). Presently Caral*luma* is gaining much importance from researchers because it possesses an array of immunostimulating activities due to the presence of various phytochemicals.

Caralluma Adscendens

Caralluma adscendens var. *gracilis* was found to contain a new pregnane glycoside (Figure 1) in diethyl ether fraction of ethanolic extract along with another known pregnane glycoside (Reddy *et al.*, 2011).

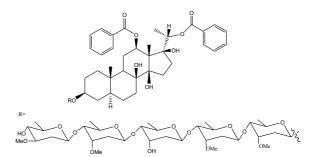


Figure 1: Pregnane glycoside from *Caralluma* adscendens var. gracilis

Caralluma Attenuata

A pentacyclic triterpenoid (Figure 2) was isolated from the n-hexane extract of roots of Caralluma *attenuata* (Jayalakshmi *et al.*, 2016). Phytochemical investigation of *Caralluma attenuata root* extracts (butanone, ethylene acetate and n-butanol extracts) has shown the presence of saponins, flavonoids or flavonoid glycosides and steroids/triterpenoids in *Caralluma attenuata* and absence of cardiac glycosides and alkaloids (Kiranmayee *et al.*, 2015).

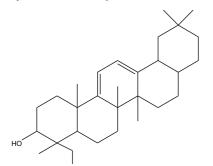


Figure 2: Pentacyclic triterpenoid from *caralluma attenuate*

Caralluma Dalzielli

Five new steroidal glycosides (Figure 3) (Caradalzieloside A-E) were isolated from CHCl₃/MeOH extract of *Caralluma dalzielii* by Oyama *et al.* (2007).

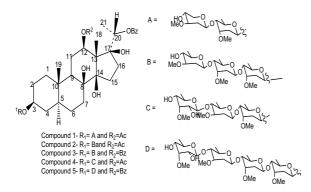


Figure 3: Steroidal glycosides from *Caralluma* dalzielii

New pregnane glycosides (27 compounds) (Figure 4) were isolated from a methanolic extract of *Caralluma dalzielii* (De Leo *et al.,* 2005). High cytotoxic activities were shown by most of the extracts.

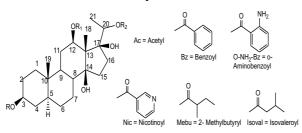


Figure 4: New pregnane glycosides from Caralluma dalzielii

Caralluma Fimbriata

Six new pregnane glycosides (2-7) (Figure 5.a-c) and their aglycon were isolated from ether fraction of ethanolic extract of *Caralluma adscendens var. Fimbriata*. Also, another six pregnane glycosides (8-13) were isolated from the butanolic fraction (Kunert *et al.*, 2008).

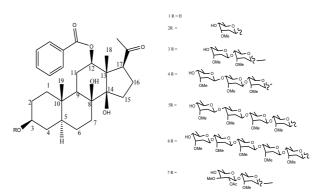


Figure 5a: Pregnane glycosides (5-7) from Caralluma adscendens var. fimbriata

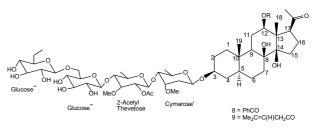


Figure 5b: Pregnane glycosides (8 & 9) from *C. adscendens var. fimbriata*

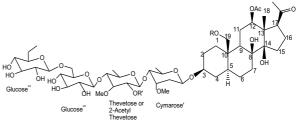


Figure 5c: Pregnane glycosides (10-13) from *C. adscendens var. fimbriata*

Oleic acid (21.08%) and n-hexadecanoic acid (44.23%) were isolated from a methanolic extract of *Caralluma Fimbriata* Wall through GC-MS analysis (Priya *et al.*, 2011).

Caralluma Flava (Desmidorchis Flava)

Four tetrasaccharide pregnane glycosides (desflavasides A-D) (Figure 6) were identified and characterized from the sap of *Desmidorchis Flava* (Raees *et al.*, 2016).

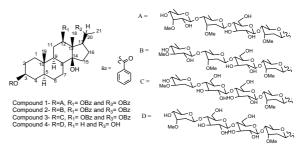


Figure 6: Tetrasaccharide pregnane glycosides (desflavasides A-D) from *Caralluma flava*

A new pregnane glycoside (nizwaside) (Figure 7) was isolated from the sap of *Desmidorchis flava* and

it was proved as an effective anticancer agent against breast cancer cells like MDA, MB231compared to known cancer drug Doxorubicin (Hussain *et al.*, 2015). However, the antioxidant activity of nizwaside was found to be weak.

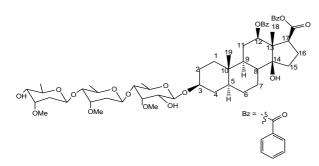


Figure 7: Pregnane glycoside (nizwaside) from *Caralluma flava*

Two new pregnane glycosides (desmiflavasides A and B) (Figure 8) were isolated from the sap of *Desmidorchis flava* and desmiflavasides B proved to contain anti-proliferation activity against breast cancer cells at a concentration of 100 mg/mL and 75 mg/mL. On the other hand, urease enzyme inhibition, as well as antioxidant activities for them, were found to be weak (Raees *et al.*, 2015).

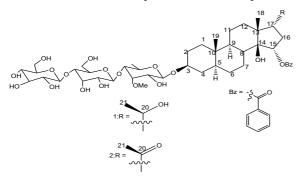


Figure 8: Pregnane glycosides (desmiflavasides A and B) from *Caralluma flava*

Caralluma Indica/Boucerosia Indica

Kunert *et al.* (2006) (Figure 9a & b) isolated novel steroidal glycosides *Caralluma* species. Stalagmoside I –V (1-5) are isolated from butanol fraction of *Caralluma stalagmifera* and indicoside I and II (7 & 8) are isolated from the ether fraction of *Caralluma indica.*

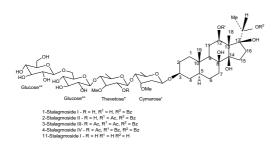


Figure 9a: Stalagmoside I-IV from Caralluma stalagmifera

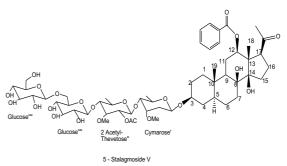
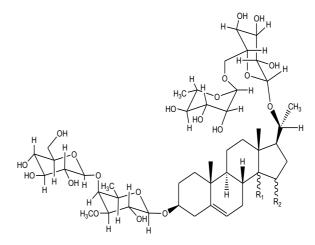


Figure 9b: Stalagmoside V from Caralluma stalagmifera

Caralluma Lasiantha/Boucerosia Lasiantha

Caralluma lasiantha (syn. Boucerosia lasiantha) belongs to the family Asclepiadaceae and its local name is Kundeti Kommulu (in Telugu)/Sirumankeerai (in Tamil) (Arinathan et al., 2007). Caralluma lasiantha is succulent inhabit and is used as an indoor ornamental plant (Reddy et al., 2012). It grows wild in Anantapur, Chittoor and surrounding places of Andhra Pradesh, India. To reduce the body heat, fresh rootless Caralluma lasiantha (a dose of 10 g, twice a day for three days) is used in India (Vikneshwaran et al., 2008). Malladi et al. (2017) reviewed the role of *Caralluma lasiantha* in traditional Indian medicine. Two new bisdesmosidic C₂₁ pregnane steroidal glycosides (lasianthosides-A and B) (Figure 10) were isolated from the n-butanol fraction of ethanolic extract of Caralluma lasiantha (Qiu et al., 1999). A flavones glycoside (luteolin-4-O-neohesperiodoside) (Figure 11) was isolated from a methanolic extract of Caralluma lasiantha (Ramesh et al., 1999b). Stigmasterol and C₂₁ Pregnane Steroid (3β,14βdihydroxy-14β-pregn-5-en-20-one) were isolated from the less polar solvent extracts (Malladi et al., 2017 a,b).



Lasianthoside-A –R₁, R₂ = Δ^{14-15} , Lasianthoside-B – R₁ = β -OH, R₂ = H

Figure 10: Lasianthosides A and B from Caralluma lasiantha

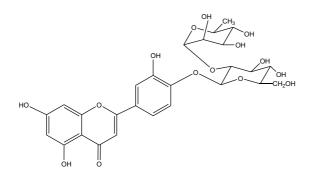


Figure 11: Luteoline-4-O-neohesperiodoside from *Caralluma lasiantha*

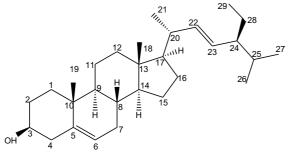


Figure 12: Stigmasterol

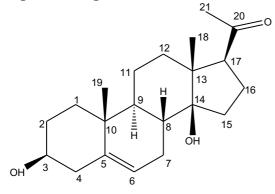


Figure 13: 3β , 14β -dihydroxy- 14β -pregn-5-en-20-one

Caralluma Negevensis

Two megastigmane glycosides (1 & 2) (Figure 14), as well as two new flavone glycosides (3 & 4), were isolated from a methanolic extract of *Caralluma negevensis* (Bader *et al.*, 2003).

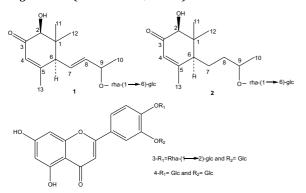
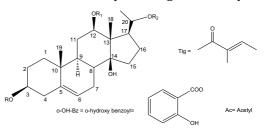
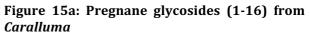


Figure 14: Megastigmane glycosides (1 & 2) from *Caralluma negevensis Caralluma Negevensis*

New pregnane glycosides (1-20) (Figure 15a and b) were isolated from chloroform and methanol extracts of *Caralluma negevensis* (Braca *et al.*, 2002). Compounds 1-16 were found to contain bouncer in aglycon moiety, compounds 17-19 were shown to contain the calogenin structure and 20^{th} compound contain 5α -dihydrocalogenin moiety.





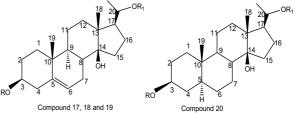


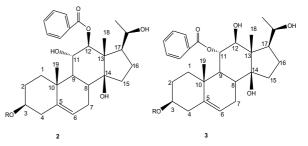
Figure 15b: Pregnane glycosides (17-20) from *Caralluma negevensis*

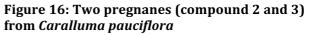
Caralluma Nilagiriana

Phytochemicals of *Caralluma nilagiriana* was evaluated by using UV-VIS and FTIR spectroscopic techniques (Renuka *et al.*, 2016). GC-MS analysis on ethanolic extract of *Caralluma nilagiriana* shows the presence of active components like alkaloids, flavonoids, phenolic compounds, steroids, terpenoids and tannins (Ranganathan *et al.*, 2014). Rutin, a bioflavonoid present in the ethanolic extract of shoots of *Caralluma nilagiriana* was determined by high-performance thin layer chromatography (HPTLC) (Renuka *et al.*, 2014).

Caralluma Pauciflora

Two pregnanes (Figure 16) were isolated from diethyl ether fraction of ethanolic extract of *Caralluma pauciflora* along with known compounds like carumbelloside-III and dihydro russelioside (Reddy *et al.*, 2011).





Caralluma Penicillata

Four acylated pregnane glycosides (Figure 17) were identified from the methanolic extract of *Caralluma penicillata* (Abdallah *et al.*, 2013).

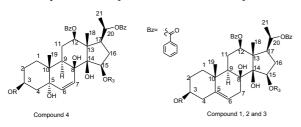


Figure 17: Four acylated pregnane glycosides from *Caralluma penicillata*

C₁₅ oxypregnane glycosides (penicillosides A–C) (Figure 18) were isolated from chloroform fraction of ethanol extract of *Caralluma penicillata* (Abdel-Sattar *et al.*, 2001).

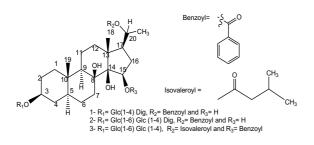


Figure 18: C₁₅ oxypregnane glycosides (penicillosides A-C) from *Caralluma penicillata*

Caralluma Quadrangular

Russelioside B (Figure 19), a pregnane glycoside isolated from the n-butanol fraction of methanol extract of *Caralluma quadrangular* was recommended for managing of diabetes based on its potent antihyperlipidemic effect (Abdel-Sattar *et al.*, 2016).

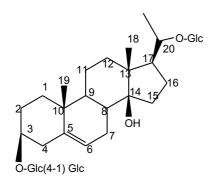


Figure 19: Russelioside B from Caralluma quadrangular

Caralluma Retrospiciens

A polyoxy pregnane glycoside (retrospinoside 1) (Figure 20) was identified from the n-butanol fraction of methanol extract *Caralluma retrospections* (Elsebai and Mohamed, 2015).

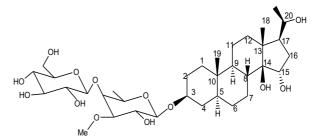


Figure 20: Polyoxy pregnane glycoside (retrospinoside 1) from *Caralluma retrospiciens*

Six polyoxy pregnane glycosides (Figure 21a, b) were extracted from ether extracts of *Caralluma retrospiciens* and all of them exhibited cytotoxic property but compound-2 shown high cytotoxic activity (Halaweish *et al.*, 2004).

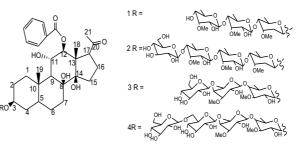


Figure 21a: Polyoxy pregnane glycosides (1-4) from *Caralluma retrospiciens*

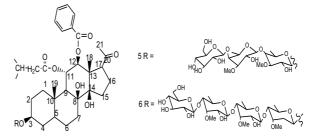


Figure 21b: Polyoxy pregnane glycosides (1-4) from *Caralluma retrospiciens*

Caralluma Russeliana

Two triterpenoids (3 and 4), two sterols (5 and 6) and two pregnane glycosides (1 and 2) (Figure 22.a-c) were isolated from the chloroform extract of *Caralluma russeliana* (Abdel-Mogib and Raghib, 2013).

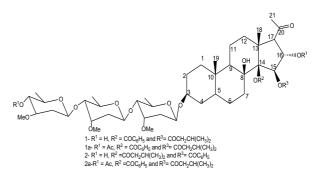


Figure 22a: Pregnane glycosides (1 and 2) from *Caralluma russeliana*

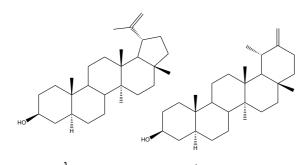


Figure 22b: Pregnane glycosides (1 and 2) from *Caralluma russeliana*

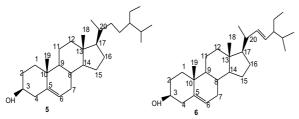


Figure 22c: Sterols (5 and 6) from *Caralluma russeliana*

Four acylated pregnane glycosides (russeliosides E–H) (Figure 23) were extracted from chloroform extracts of *Caralluma russeliana* (Abdel-Sattar *et al.,* 2007).

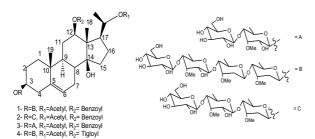


Figure 23: Acylated pregnane glycosides (russeliosides E-H) from *Caralluma russeliana*

A known flavone glycoside and four pregnane glycosides (Figure 24) were extracted from *Caralluma russeliana* in its n-butanol fraction of ethanol extract (Abdul-Aziz Al-Yahya *et al.*, 2000).

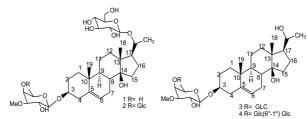


Figure 24: Pregnane glycosides (russeliosides E-H) from *Caralluma russeliana*

Caralluma Sinaica

Caralluma sinaica yielded six pregnane glycosides along with seven pregnanes and three flavonoids (Figure 25) from its chloroform extract (Al-Massarani *et al.*, 2012). Quinone reductase induction was evaluated for all these isolated compounds.

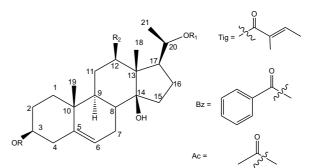


Figure 25: Pregnane glycosides, pregnanes and flavonoids from *Caralluma sinaica*

Boucerosia Truncato Coronato/Caralluma Truncato-Coronato

Phytochemical constituents present in ethanolic extracts of *Caralluma truncato coronato* were studied by GC-MS analysis by Kalimuthu *et al.* (2013). The results obtained have shown the presence of furan, 2-butyltetrahydro- (0.57%), β -sitosterol (1.89%), acetic acid, 5-(dimethyle-6-oxocyclohexylidene)-3-methyl-pent-3-enyl ester (3.78%), β -tocopherol (4.91%), squalene and lupeol (5.67%), vitamin E (9.45%) and thunbergol (68.05%).

Caralluma Tuberculata

Two steroidal glycosides (pregnane glycoside and androstan glycoside) (Figure 26) were found in ethyl acetate extract of *Caralluma tuberculata* and exhibited moderate cytotoxic activity against three human breast cancer cell lines (Waheed *et al.*, 2011).

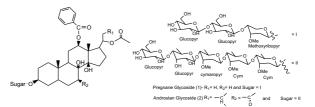


Figure 26: Steroidal glycosides from *Caralluma tuberculata*

Five pregnane glycosides (caratuberside A-E) and preganan glycoside-russelioside (Figure 27) were isolated from chloroform fraction of MeOH extract of *Caralluma tuberculata*. The isolated pregnane glycosides exhibited antimalarial, antitrypanosomal and cytotoxic potentials (Abdel-Sattar *et al.*, 2008).

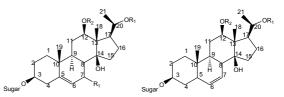


Figure 27: Pregnane glycosides (caratuberside A-E) from *Caralluma tuberculata*

Caralluma Umbellata

Kalyani *et al.* (2013) (Figure 28) isolated a novel pregnane glycoside from ether eluates of methanol and benzene fractions of ethanolic extract of roots of *Caralluma umbellata*.

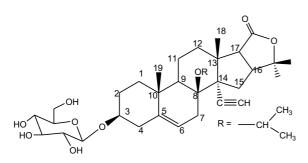


Figure 28: Pregnane glycoside from *Caralluma umbellata*

Presence of thirty-two compounds was found in a phytochemical analysis of leaves of *Caralluma umbellata* by Jerome Jeyakumar *et al.* (2013). The extracted phytochemical compounds include terpenoids, flavonoids, tannins, phenols, glycosides sterols and saponins. Two new pregnane compounds (CRUR I and CRUR II) (Figure 27) were isolated from the roots of *Caralluma umbellata* (Kishore *et al.*, 2010).

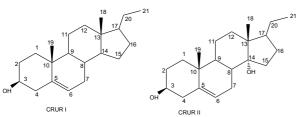


Figure 29: Pregnane compounds from *Caralluma umbellate*

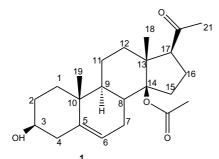


Figure 30: Pregnane steroid from *Caralluma umbellate*

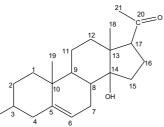


Figure 31: C₂₁ steroid from *Caralluma umbellata*

Steroidal glycosides were found in different solvent fractions of ethanol extract of *Caralluma umbellata*. The isolated steroidal glycosides are bourcergenin (new pregnene type steroid) from toluene fraction, carumbelloside-VI (bisdesmosidic glycoside) from n-butanol fraction, and carumbellosides I–IV (new tetroxides of boucergenin) from toluene fraction (Kunert *et al.*, 2009). A new pregnane steroid (Figure 28) containing formyl group was isolated and characterized from *Caralluma umbellata* stems along with three known steroidal compounds (Babu *et al.*, 2008).

A C₂₁ steroid (Figure 31) was isolated from the toluene fraction of ethanol extract of *Caralluma umbellata* from the whole plant (Ramesh et al., 2005).

Caralluma Wissmannii

Two pregnane glycosides (5, 6) along with five known compounds (viz., β -sitosterol, stigmasterol, luteolin 3',4'-di-O- β -D-glucopyranoside, 3,4-sec-up-20-(29)-en-3-oic acid methyl ester and lupeol) (Figure 30.a and b) were isolated in the phytochemical investigation on *Caralluma wissmannii* (Dawidar *et al.*, 2012).

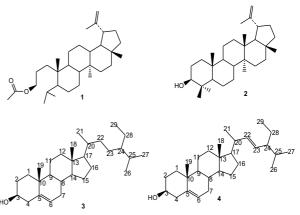


Figure 32a: (1) 3,4-seco-lup-20(29)-en-3-oic acid methyl ester, (2) lupeol (3) β-sitosterol and (4) stigmasterol from *Caralluma wissmannii*

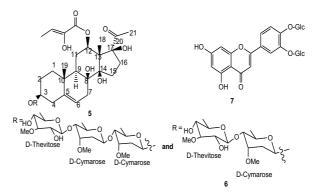


Figure 32b: (5, 6) Pregnane glycosides and (7) Luteolin 3',4'-di-O-β-D-glucopyranoside from *Caralluma wissmannii*

Name of the phytochemical	als isolated from different plants of th Name of the plant	Reference
Flavone glycoside,	Caralluma arabica	Kamil et al., 1999
Luteoline-4'-0-neohesperiodo-	Caralluma umbellata	Ramesh et al., 1999b
side	Caralluma lasiantha	Ramesh et al., 1999b
	Caralluma attenuata	Ramesh et al., 1998, 1999b
	Caralluma tuberculata	Rizwani et al., 1990
Flavones glycosides and	Caralluma negevensis	Bader et al., 2003
megastigmane glycosides	Saranana negovensis	Bader et all, 2000
Pregnane glycosides	Caralluma quadrangula	Abdel-Sattar et al., 2016
	Caralluma flava (Desmidorchis flava)	Raees et al., 2015, 2016
	Caralluma flava (Desmidorchis flava)	Hussain et al., 2015
	Caralluma retrospiciens	Elsebai and Mohamed, 2015
	Caralluma umbellata	Kalyani et al., 2013
	Caralluma wissmannii	Dawidar et al., 2012
	Caralluma adscendens var. gracilis	Reddy et al., 2011
	Caralluma adscendens var.fimbriata	Kunert et al., 2008
	Caralluma dalzielli	De Leo et al., 2005
	Caralluma negevensis	Braca et al., 2002
	Caralluma russeliana	Al-Yahya et al., 2000
	Caralluma retrospiciens	Halim and Khalil., 1996
	Caralluma umbellata	Lee-Juian et al., 1994
	Boucerosia aucheriana	Tanaka et al., 1990
	Boucerosia aucheriana	Hayashi et al., 1988
	Caralluma tuberculata	Ahmad et al., 1988
Acylated Pregnane glycosides	Caralluma quadrangula	Abdallah et al., 2013
neyhteet regnane grycosides	Caralluma sinaica	Al-Massarani et al., 2012
	Caralluma tuberculata	Abdel-Sattar et al., 2008
	Caralluma russeliana	Abdel-Sattar et al., 2007
Pregnane glycoside diesters	Caralluma russeliana	Abdel-Mogib and Raghib,
	Sar anama rabbenana	2013
Oxy Pregnane glycosides	Caralluma pencillata	Abdel-Sattar et al., 2002
	Caralluma pencillata	Abdel-Sattar et al., 2001
Polooxy Pregnane glycosides	Caralluma retrospiciens	Halaweish et al., 2004
Steroidal glycosides	Caralluma umbellata	Kunert et al., 2009
	Caralluma stalagmifera	Kunert et al., 2006
	Caralluma dalzielii	Oyama et al., 2007
	Caralluma lasiantha	Qiu et al., 1999
	Caralluma umbellata	Qiu et al., 1997
	Caralluma indica	Kunert et al., 2006
Acylated steroidal glycosides	Caralluma tuberculata	Waheed et al., 2011
Pregnane steroids/pregnanes	Caralluma umbellata	Ramesh et al., 2005
r ognane ster ords/ prognanes	Caralluma pauciflora	Reddy et al., 2011
	Caralluma umbellata	Kishore et al., 2010
	Caralluma umbellata	Babu et al., 2008
Pregnane ester aglycones	Caralluma retrospiciens	Khalil, 1995
Triterpenoid	Caralluma attenuata	Jayalakshmi et al., 2016
Terpenoids	Caralluma europaea	Formisano et al., 2009
β-sitosterol, lupeol	Caralluma wissmannii	Dawidar et al., 2012
p-situsteroi, iupeoi	Caralluma adscendens var. gracilis	Reddy et al., 2012
	sai anama aaseenaens vari graems	1.000 00 00 00 1 1
	Caralluma truncato-coronata	Kalimuthu et al., 2013

Table 1: A list of phytochemicals isolated from different plants of the Caralluma genus

In continuation to the above list of phytochemicals collection, efforts of various researchers on their study pertaining to different species of *Caralluma* are compiled in Table-1.

CONCLUSION

Caralluma is a genus used as traditional medicine and is a potential source for phytochemicals with medicinal uses. Thorough literature collection shows few species of *Caralluma* are explored for isolation of phytochemicals from them. Enthusiastic researchers in this field can extend their investigation for further exploration of many species (like Caralluma acutanaula. other Caralluma Arabica, Caralluma cicatricose, Caralluma edulis, Caralluma Europaea, Caralluma *longidens*) for phytochemicals present in them.

REFERENCES

A. Waheed, J. Barker, S.J. Barton, G.M. Khan, Q. Najm-us-Saqib, M. Hussain, S. Ahmed, C. Owen, M.A. Carew, Novel acylated steroidal glycosides from *Caralluma tuberculata* induce caspase-dependent apoptosis in cancer cells, Journal of Ethnopharmacol, 137(3), 2011pp. 1189-1196.

Abdallah HM, Osman AM, Almehdar H, Abdel-Sattar E. Acylated pregnane glycosides from *Caralluma quadrangular*, Phytochemistry, 88, 2013pp. 54-60.

- Abdel-Mogib M, Raghib HM. Two new pregnane glycoside diesters from *Caralluma russeliana*, Natural Product Research, 27(14), 2013pp.1287-1292.
- Abdel-Sattar E, Ahmed AA, Hegazy ME, Farag MA, Al-Yahya MA. Acylated pregnane glycosides from *Caralluma russeliana*, Phytochemistry, 68(10), 2007pp. 1459-63.

Abdel-Sattar E, Al-Yahya MA, Nakamura N, Hattori M. Penicillosides A–C, C-15 oxypregnane glycosides from *Caralluma penicillata*, Phytochemistry, 57(8), 2001 pp. 1213-1217.

Abdel-Sattar E, EL-Maraghy SA, El-Dine RS, Rizk SM. Russelioside B, a pregnane glycoside ameliorates hyperglycemia in streptozotocin induced diabetic rats by regulating key enzymes of glucose metabolism, Chemico-Biological Interactions, 252, 2016pp. 47-53.

Abdel-Sattar E, Meselhy MR, Al-Yahya MA. New oxypregnane glycosides from *Caralluma penicillata*, Planta medica, 68(05), 2002pp. 430-434.

Abdel-Sattar E. Harraz FM, Al-Ansari SM, El-Mekkawy S, Ichino C, Kiyohara H, Ishiyama A, Otoguro K, Omura S, Yamada H. Acylated pregnane glycosides from *Caralluma tuberculata* and their antiparasitic activity, Phytochemistry, 69(11), 2008pp. 2180-2186.

Abdul-Aziz Al-Yahya M, Abdel-Sattar E, Guittet E. Pregnane glycosides from *Caralluma russeliana*, Journal of Natural Products, 63(10), 2000 pp. 1451-1453.

Ahmad MM, Shaikh MM. Improvement in glucose tolerance by *Caralluma tuberculata* Acacia

nilotica and Papaver somniferum, Pakistan Journal Zoology, 1989.

Ahmad VU, Usmanghani K, Rizwani GH. New pregnane glycosides from *Caralluma tuberculata*, Journal of Natural Products, 51(6), 1988pp. 1092-1097.

Al-Massarani SM, Bertrand S, Nievergelt A, El-Shafae AM, Al-Howiriny TA, Al-Musayeib NM, Cuendet M, Wolfender JL. Acylated pregnane glycosides from *Caralluma sinaica*, Phytochemistry, 79, 2012, pp. 129-140.

Arinathan V, Mohan VR, Britto A, Murugan C. Wild edibles used by Palliyars of the Western Ghats, Tamil Nadu, Indian Journal of Traditional Knowledge, 6(1), 2007pp. 163-168.

Babu KS, Malladi S, Nadh RV, Rambabu SS. Evaluation of in Vitro Antibacterial Activity of *Caralluma umbellata* Haw Used in Traditional Medicine by Indian Tribes, Annual Research & Review in Biology, 4(6), 2014pp.840.

Bader A, Braca A, De Tommasi N, Morelli I. Further constituents from Caralluma negevensis. Phytochemistry, 62(8), 2003 pp. 1277-1281.

Baker DD, Chu M, Oza U, Rajgarhia V. The value of natural products to future pharmaceutical discovery, Natural Product Reports, 24(6), 2007pp. 1225-1244.

Bhanot A, Sharma R, Noolvi MN. Natural sources as potential anti-cancer agents: A review, International Journal of Phytomedicine, 3(1), 2011pp. 09-26.

Braca A, Bader A, Morelli I, Scarpato R, Turchi G, Pizza C, De Tommasi N. New pregnane glycosides from Caralluma negevensis, Tetrahedron, 58(29), 2002pp. 5837-5848.

Chemat F, Strube J. (ed.) Green extraction of natural products: Theory and practice, John Wiley & Sons, 2015.

Costa MA, Xia ZQ, Davin LB, Lewis NG. Lewis, chapter four: Toward engineering the metabolic pathways of cancer-preventing lignans in cereal grains and other crops. In recent advances in phytochemistry, phytochemicals in human health protection, Nutrition and plant defence, ed. JT Romeo, New York, 33, 1999pp. 67-87.

Cragg DJ, Kingston GM, Newman DG, Anticancer agents from natural products, CRC press, 2011.

Dawidar AM, Mubarak AY, Abdel-Mogib M, Abdelsattar E. Phytochemical investigation of *Caralluma wissmannii*, Research Journal of Pharmaceutical, Biological and Chemical Sciences, 3, 2012pp. 882-892.

- De Leo M, De Tommasi N, Sanogo R. G, Marzocco S, Pizza C, Morelli I, Braca A. New pregnane glycosides from *Caralluma dalzielii*, Steroids, 70, 2005pp. 573-585.
- Doughari JH, Human IS, Bennade S, Ndakidemi PA. Phytochemicals as chemotherapeutic agents and antioxidants: Possible solution to the control of antibiotic-resistant verocytotoxin producing bacteria, Jornal of Medical Plants Research, 3(11), 2009pp. 839-848.
- Doughari JH, Obidah JS. Antibacterial potentials of stem bark extracts of Leptadenia lancifolia against some pathogenic bacteria, Pharmacologyonline, 3, 2008pp. 172-180.
- Elsebai MF, Mohamed IE. New pregnane glycoside derivative from *Caralluma retrospiciens* (Ehrenb), Natural product research, 29(15), 2015pp. 1426-1431.
- Formisano C, Senatore F, Della Porta G, Scognamiglio M, Bruno M, Maggio A, Rosselli S, Zito P, Sajeva M. Headspace volatile composition of the flowers of *Caralluma europaea* NE Br.(Apocynaceae). Molecules, 14(11), 2009pp. 4597-4613.
- Gandhi R. *Carallumas* of the Indian subcontinent, Indian society of cactii and succulents, New Delhi: Ram Gandhi;1999.
- Gilbert MG, A review of *Caralluma* R. Br. and its segregates, Bradleya,8,1990pp.1-32.
- Halaweish FT, Huntimer E, Khalil AT. Polyoxypregnane glycosides from *Caralluma retrospiciens*, Phytochemical Analysis, 15(3), 2004pp. 189-194.
- Halim AF, Khalil AT. Pregnane glycosides from *Caralluma retrospiciens*, Phytochemistry, 42(4), 1996pp. 1135-1139.
- Harborne JR, Introduction to ecological biochemistry, 4th edition., Academic Press, London: Elsevier,1993, 1–32.
- Hayashi K, Iida I, Nakao Y, Nakao Y, Kaneko K. Four pregnane glycosides, boucerosides AI, AII, BI and BII, from *Boucerosia aucheriana*, Phytochemistry, 27(12), 1988pp.3919-3924.
- Hussain H, Raees MA, Rehman NU, Al-Rawahi A, Csuk R, Khan HY, Abbas G, Al-Broumi MA, Green IR, Elyassi A, Mahmood T. Nizwaside: a new anticancer pregnane glycoside from the sap of *Desmidorchis Flava*, Archives of Pharmacal Research, 38(12), 2015pp. 2137-2142.
- Jayalakshmi G, Anuradha V, Ratnakumari S, Kalyani K, Babu SS. A novel pentacyclic triterpenoid

isolated from *Caralluma attenuate* root, European Journal of Pharmaceutical and Medical Research, 36, 2016pp. 342-344.

- Jerome Jeyakumar J, Kamaraj M, Nandagopalan V, Anburaja V, Thiruvengadam M. Study of phytochemical constituents in Caralluma umbellata By GG-MS Analysis, International Journal of Pharmaceutical Science Invention, 2(4), 2013pp. 37-41.
- Kalimuthu K, Prabakaran R, Kalaiyarasi, K, Jayaraman S, Sasikala T. GC-MS analysis of bioactive constituents of *Caralluma truncato-coronata* (sedgw.) gravely & mayur.(Asclepiadaceae), Asian Pacific Journal of Research, 1(9), 2013pp. 42-50.
- Kalyani K, Anuradha V, Jayalakshmi G. A novel pregnane glycoside from the *Caralluma umbellata* Haw (Asclepiadaceae) Roots, Journal of Natural Sciences Research, 3(3), 2013pp. 55-61.
- Kamil M, Jayaraj AF, Ahmad F, Gunasekhar C, Samuel S, Chan K, Habibullah M. Identification and quantification of flavonoids from *Caralluma arabica* and its quality control studies. Journal of Pharmacy and Pharmacology, 51, 1999, 225-225.
- Khalil AT. Pregnane esters from *Caralluma retrospiciens*, Fitoterapia, 66(3), 1995pp. 261-264.
- Kiranmayee P, Anitha. K, Usha R. Phytochemical Investigation of *Caralluma attenuata* (Wight) Roots, International Journal of Pharmacognosy and Phytochemical Research, 7, 2015pp. 1120-1024.
- Kishore M, Surendrababu K, Hanumantharao Y, Bindu G, Janardhan M. Chemical examination of medicinal plant *Caralluma umbellata* (Asclepiadaceae) roots, International Journal of Applied Biological and Pharmaceutical Technology, 2010pp. 545-549.
- Kunert O, Rao BV, Babu GS, Padmavathi M, Kumar BR, Alex RM, Schühly W, Simic N, Kühnelt D, Rao AV. Novel steroidal glycosides from two Indian *Caralluma* species, *Caralluma* stalagmifera and *Caralluma* indica, Helvetica Chimica Acta, 89(2), 2006pp. 201-209.
- Kunert O, Rao VG, Babu GS, Sujatha P, Sivagamy M, Anuradha S, Rao BV, Kumar BR, Alex RM, Schühly W, Kühnelt D. Pregnane glycosides from *Caralluma adscendens* var. *Fimbriata*, Chemistry & Biodiversity, 5(2), 2008pp. 239-250.
- Kunert O, Simic N. Ravinder E. Rao BV, Kumar BR, Alex RM, Kuehnelt D, Rao AV, Steroidal glycosides from *Caralluma umbellata*, Phytochemistry Letters, 2(3), 2009pp. 134-138.

- Lee-Juian L, Long-Ze L, Gil RR, Cordelly GA, Ramesh M, Srilatha B, Reddy B, Rao AV. Pregnane glycosides from *Caralluma umbellata*, Phytochemistry, 35(6), 1994pp. 1549-1553.
- Malladi S, Ratnakaram VN, Babu KS, Pullaiah T. *Caralluma lasiantha:* A review on it is vital role in Indian Traditional Medicine, Research Journal of Pharmaceutical, Biological and Chemical Sciences, 8(1), 2017pp. 873-879.
- Malladi S, Ratnakaram VN, Babu KS, Pullaiah T. Evaluation of in vitro antibacterial activity of Caralluma lasiantha for scientific validation of traditional Indian medicine, Cogent Chemistry, 3(1), 2017,1374821.
- Malladi S, Ratnakaram VN, Babu KS, Sreenivasulu M, Pharmacological review of *Caralluma R.Br*: A potential herbal genus, communicated to Asian Journal of Pharmaceutical Sciences.
- Malladi S, Ratnakaram VN, Pullaiah T. Phytochemical Screening of *Caralluma lasiantha*: Isolation of C21 Pregnane Steroid, Oriental Journal of Chemistry, 33(2), 2017pp. 963-967.
- Malladi S, Ratnakaram VN, Suresh Babu K. Phytochemical investigation of *Caralluma lasiantha*: Isolation of Stigmasterol, an active immunomodulatory agent, International Journal of Chemical Sciences, 15(1), 2017pp. 399-407.
- Mathai K, Nutrition in the adult years. Krause's Food, Nutrition, and Diet Therapy, 10th edition., (ed.) Mahan LK and Escott-Stump S, 271, 2000, 274-275.
- Meve U, Liede S. Subtribal division of Ceropegieae (Apocynaceae-Asclepiadoideae), Taxon, 53(1), 2004pp.61-72.
- Newman DJ, Cragg GM, Snader KM. Natural products as sources of new drugs over the period 1981–2002, Journal of Natural Products, 66(7), 2003pp. 1022-1037.
- Newman DJ, Cragg GM. Natural products as sources of new drugs over the 30 years from 1981 to 2010, Journal of Natural Products, 75(3), 2012pp. 311-335.
- Nweze EI, Okafor JI, Njoku O. Antimicrobial activities of methanolic extracts of Trema guineensis (Schumm and Thorn) and Morinda Lucida benth used in Nigerian, Bio-research, 2(1), 2004pp. 39-46.
- Oyama M, Iliya I, Tanaka T, Iinuma M. Five new steroidal glycosides from *Caralluma dalzielii*, Helvetica Chimica Acta, 90(1), 2007pp. 63-71.
- Priya D, Rajaram K, Suresh Kumar P. Phytochemical studies and GC-MS analysis of *Caralluma Fim*-

briata wall. International Journal of Pharmaceutical Research and Development, 3(10), 2011pp. 105-110.

- Qiu SX, Cordell GA, Kumar BR, Rao YN, Ramesh M, Kokate C, Rao AV. bisdesmosidic pregnane glycosides from Caralluma lasiantha, Phytochemistry, 50(3), 1999 pp. 485-491.
- Qiu SX, Lin LZ, Cordell GA, Ramesh M, Kumar BR, Radhakrishna M, Mohan GK, Reddy BM, Rao YN, Srinivas B, Thomas NS. Acylated C-21 steroidal bisdesmosidic glycosides from *Caralluma umbellata*, Phytochemistry, 46(2), 1997pp. 333-340.
- Raees MA, Hussain H, Al-Rawahi A, Csuk R, Al-Ghafri A, Rehman NU, Elyassi A, Green IR, Mahmood T, Al-Harrasi A. Desflavasides AD: Four new tetrasaccharide pregnane glycosides from *Desmidorchis flava*, Phytochemistry Letters, 16, 2016pp. 230-235.
- Raees MA, Hussain H, Rehman NU, Khan HY, Abbas G, Al-Rawahi A, Elyassi A, Al-Amri IS, Green IR, Al-Broumi MA, Mahmood T. Desmiflavasides A and B: Two new bioactive pregnane glycosides from the sap of *Desmidorchis Flava*, Phytochemistry Letters, 12, 2015pp. 153-157.
- Ramesh M, Rao YN, Kumar MR, Mohan GK, Kumar BR, Rao AA, Krishna MR, Reddy BM. Flavone glycoside from three *Caralluma* species, Biochemical Systematics and Ecology, 27(1), 1999bpp. 85-86.
- Ramesh M, Rao YN, Rao AA, Prabhakar MC, Rao CS. Muralidhar N, Reddy BM. The antinociceptive and anti-inflammatory activity of a flavonoid isolated from *Caralluma attenuata*, Journal of Ethnopharmacol, 62(1), 1998pp. 63-66.
- Ramesh M, Ravi Kumar B, Chandra K, Venkatesdh S, Appa Rao AV. C21 Pregnane Steroid from *Caralluma umbellata*, Natural Products Science, 11(2), 2005pp. 115-117.
- Ranganathan, D. Phytochemical analysis of Caralluma nilagiriana using GC-MS. Journal of Pharmacognosy and Phytochemistry, 3(1), 2014pp. 155-159.
- Reddy KD, Rao BV, Babu GS, Kumar BR, Braca A, Vassallo A, De Tommasi N, Rao GV, Rao AV. Minor pregnanes from *Caralluma adscendens* var. *gracilis* and *Caralluma pauciflora*. Fitoterapia, 82(7), 2011pp. 1039-1043.
- Reddy SR, Reddy AM, Yasodamma N. Exploration of the wild ornamental flora of YSR District, Andhra Pradesh, India, Indian Journal of Fundamental and Applied Life Sciences, 2(1), 2012pp. 192-199.

- Renuka B, Sanjeev B, Ranganathan D. Evaluation of phytoconstituents of Caralluma nilagiriana by FTIR and UV-VIS spectroscopic analysis, Journal of Pharmacognosy and Phytochemistry, 5(2), 2016 pp. 105.
- Renuka, B. A high-performance thin layer chromatography determination and quantification of rutin in *Caralluma nilagiriana*, an endemic medicinal plant, Chemistry, 1(2), 2014pp. 12-22.
- Rizwani GH, Usmanghani K, Ahmad M, Ahmad VU. Flavone glycosides of *Caralluma tuberculata* NE Brown, Pakistan Journal of Pharmaceutical Sciences, 3(2), 1990pp. 27-32.
- Saganuwan A. Some medicinal plants of Arabian Peninsula, Journal of Medicinal Plants Research, 4(9), 2010pp. 767-89.
- Saxena M, Saxena J, Nema R, Singh D, Gupta A. Phytochemistry of medicinal plants, Journal of Pharmacognosy and Phytochemistry, 1(6), 2013pp. 168-182.
- Smolinski ms, Hamburg MA, Lederberg J. (ed.) Microbial threats to health: Emergence, detection, and response. Washington, DC: Institute of Medicine, National Academies Press. 2003, 203-210.
- Suresh Babu K, Rama Subba Rao V, Vyasa Radhakrishnan S, Madhusudana Rao J, Siva Rambabu S. A new pregnane steroid from the stems of *Caralluma umbellata*, Journal of Asian Natural Products Research, 10(11), 2008pp. 1013-1016.
- Tanaka T, Tsukamoto S, Hayashi K. Pregnane glycosides from Boucerosia aucheriana, Phytochemistry, 29(1), 1990pp. 229-237.
- Temidayo AR. Extraction and isolation of flavonoids present in the methanolic extract of leaves of Acanthospermum hispidium Dc, Global Journal of Medicinal Plant Research, 1(1), 2013pp. 111-123.
- Van den Bogaard AE, Stobberingh EE. Epidemiology of resistance to antibiotics: links between animals and humans, International Journal of Antimicrobial Agents, 14(4), 2000 pp. 327-335.
- Vikneshwaran D, Viji M, Rajalakshmi K. Ethnomedicinal plants survey and documentation related to Paliyar community, Ethnobotanical Leaflets, 2008(1), 2008pp. 1108-1115.